

# Species List

Humber Valley, 2017

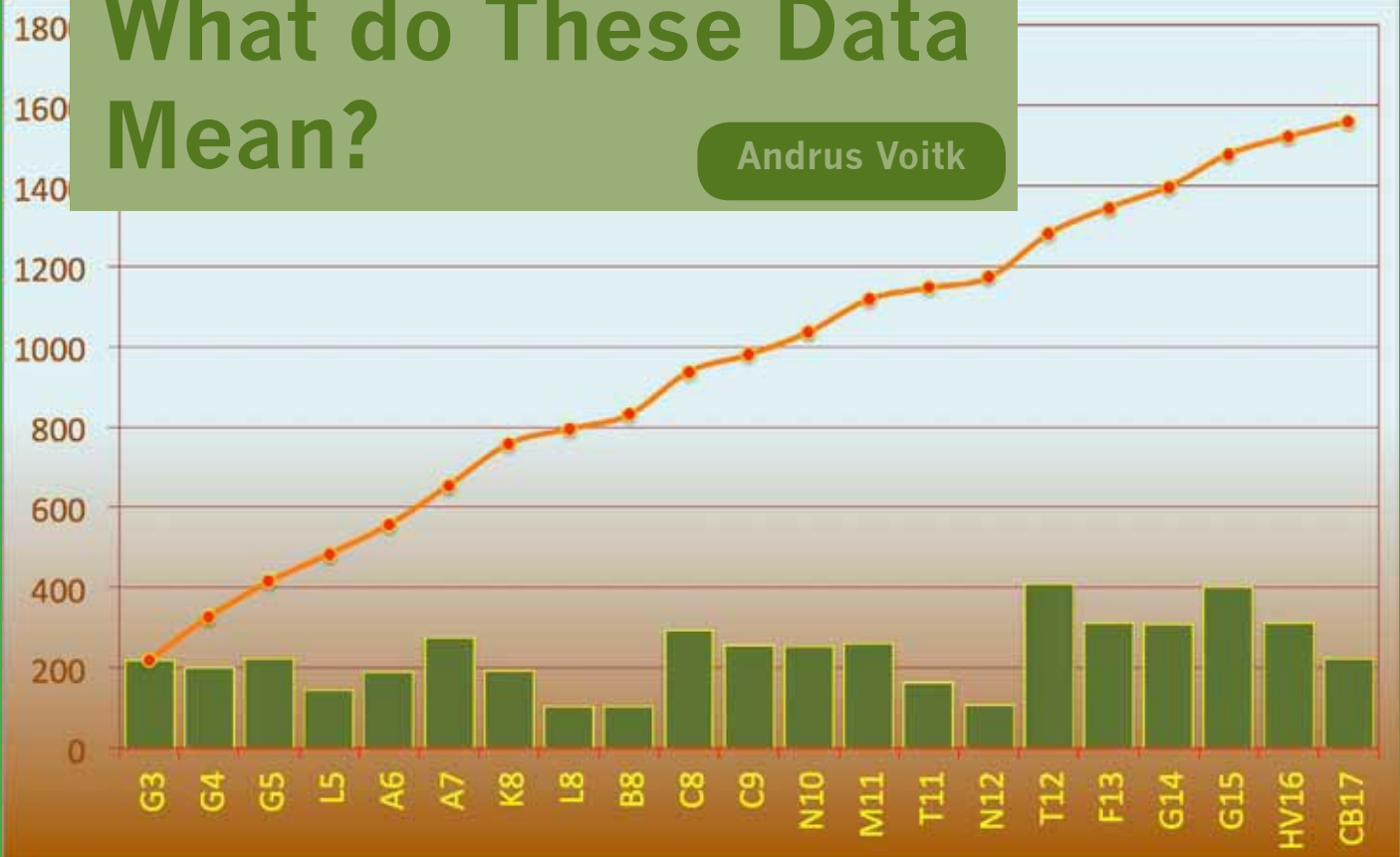


New species to the cumulative list are shown in **black boldface print**.

<i>Agaricus campestris</i>	<i>Cortinarius collinitus</i>	<i>Hygrocybe squamulosa</i>	<i>Peziza michelii</i>
<i>Amanita bisporigera</i>	<i>Cortinarius delibutus</i>	<b><i>Hymenoscyphus repandus</i></b>	<i>Peziza sedia</i>
<b><i>Amanita brunnescens</i></b>	<i>Cortinarius flexipes</i>	<i>Hypomyces chrysospermus</i>	<b><i>Phaeohelotium epiphyllum</i></b>
<i>Amanita ceciliae</i>	<i>Cortinarius grosmorenensis</i>	<i>Hypomyces hyalinus</i>	<i>Phaeolus schweinitzii</i>
<i>Amanita flavoconia</i>	<i>Cortinarius huronensis</i>	<i>Hypomyces lactifluorum</i>	<i>Phellinus chrysoloma</i>
<i>Amanita fulva</i>	<i>Cortinarius malicorius</i>	<b><i>Hypomyces papalosporae</i> var.</b>	<i>Phellinus laevigatus</i>
<i>Amanita muscaria</i> var.	<i>Cortinarius scaurus</i>	<b><i>americana</i></b>	<i>Phellinus prunicola</i>
<i>guessowii</i>	<i>Cortinarius stillatitius</i>	<i>Hypoxyton fuscum</i>	<i>Phellodon niger</i>
<i>Amanita porphyria</i>	<i>Cortinarius subtortus</i>	<i>Imleria badia</i>	<b><i>Picipes tubaeformis</i></b>
<i>Amanita rubescens</i>	<b><i>Cortinarius tofaceus</i></b>	<b><i>Inocybe comatella</i></b>	<i>Piptoporus betulinus</i>
<i>Amanita wellsii</i>	<b><i>Cortinarius venetus</i></b>	<b><i>Inocybe egenula</i></b>	<i>Plicaturopsis crispa</i>
<i>Ampulloclitocybe clavipes</i>	<i>Craterellus tubaeformis</i>	<i>Inocybe geophylla</i>	<i>Polyporus varius</i>
<b><i>Annulohyphoxylon multifforme</i></b>	<i>Crucibulum laeve</i>	<b><i>Inocybe hystrix</i></b>	<b><i>Porphyrellus fumosipes</i></b>
<i>Antrodia heteromorpha</i>	<i>Cudonia circinans</i>	<b><i>Inocybe jacobi</i></b>	<i>Postia tephroleuca</i>
<i>Aphroditeola olida</i>	<b><i>Cystolepiota seminuda</i></b>	<i>Inocybe lanuginosa</i>	<i>Pseudohydnum gelatinosum</i>
<i>Apiosporina morbosa</i>	<i>Dacrymyces chrysospermus</i>	<i>Inocybe lilacina</i>	<i>Puccinia poarum</i>
<i>Arrhenia sphagnicola</i>	<b><i>Daedaleopsis confragosa</i></b>	<i>Inonotus obliquus</i>	<i>Pycnoporellus fulgens</i>
<b><i>Aurantioporthe corni</i></b>	<b><i>Daldinia childii</i></b>	<i>Ischnoderma resinsum</i>	<i>Ramariopsis kunzei</i>
<i>Austroboletus gracilis</i>	<i>Elaphomyces muricatus</i>	<i>Jahnoporus hirtus</i>	<b><i>Ramariopsis tenuiramosa</i></b>
<i>Boletus edulis</i>	<i>Entoloma bicolor</i>	<i>Laccaria bicolor</i>	<i>Rhodocollybia maculata</i> var.
<i>Calocera viscosa</i>	<i>Entoloma formosum</i>	<i>Laccaria laccata</i> var.	<i>scorzonerea</i>
<b><i>Cantharellus amethysteus</i></b>	<i>Entoloma incanum</i>	<i>pallidifolia</i>	<b><i>Rhytisma curtisii</i></b>
<b><i>Cantharellus camphoratus</i></b>	<b><i>Entoloma jubatum</i></b>	<i>Laccaria striatula</i>	<i>Rhytisma salicinum</i>
<i>Catathelasma ventricosum</i>	<b><i>Entoloma luteum</i></b>	<i>Lachnellula agassizii</i>	<b><i>Rhytisma solidagenis</i></b>
<i>Ceratiomyxa fruticulosa</i>	<i>Entoloma quadratum</i>	<i>Lachnum virgineum</i>	<i>Rickenella fibula</i>
<i>Chalciporus piperatus</i>	<i>Entoloma sericellum</i>	<i>Lactarius camphoratus</i>	<i>Russula abietina</i>
<b><i>Chalciporus rubinellus</i></b>	<i>Entoloma serrulatum</i>	<i>Lactarius deterrimus</i>	<i>Russula brevipes</i>
<i>Chlorociboria aeruginascens</i>	<i>Entoloma strictius</i> var.	<i>Lactarius helvus</i>	<i>Russula cf betularum</i>
<i>Chlorociboria aeruginosa</i>	<i>isabellinum</i>	<i>Lactarius lignyotus</i>	<i>Russula decolorans</i>
<i>Chrysomphalina chrysophylla</i>	<i>Exobasidium vaccinii</i>	<i>Lactarius mucidus</i>	<i>Russula compacta</i>
<i>Chrysomyxa ledicola</i>	<i>Fomes fomentarius</i>	<i>Lactarius scrobiculatus</i> var.	<i>Russula dissimulans</i>
<b><i>Chrysomyxa pirolata</i></b>	<i>Fomitopsis cajanderi</i>	<i>canadensis</i>	<i>Russula fragilis</i>
<b><i>Clavaria fragilis</i></b>	<i>Fomitopsis ochracea</i>	<i>Lactarius tabidus</i>	<b><i>Russula humidicola</i></b>
<i>Clavaria rosea</i>	<i>Fomitopsis pinicola</i>	<i>Lactarius thyinos</i>	<i>Russula montana</i>
<i>Clavaria vermicularis</i>	<i>Fomitopsis rosea</i>	<i>Leccinum holopus</i>	<i>Russula paludosa</i>
<i>Clavulina cinerea</i>	<i>Fuligo septica</i>	<i>Leccinum scabrum</i>	<i>Russula peckii</i>
<i>Clavulina rugosa</i>	<i>Galerina atkinsoniana</i>	<i>Leotia lubrica</i>	<b><i>Scleroderma bovista</i></b>
<i>Clavulinopsis fusiformis</i>	<i>Ganoderma applanatum</i>	<i>Leptoporus mollis</i>	<i>Scutellinia scutellata</i>
<i>Clitopilus prunulus</i>	<i>Gliophorus irrigatus</i>	<b><i>Leucocoprinus cepistipes</i></b>	<b><i>Sebacina concrescens</i></b>
<i>Collybia cirrhata</i>	<i>Gliophorus laetus</i>	<i>Lycogala epidendrum</i>	<i>Steccherinum ochraceum</i>
<i>Conocybe apala</i>	<i>Gloeophyllum sepiarium</i>	<i>Lycoperdon perlatum</i>	<b><i>Stereum ostrea</i></b>
<i>Conocybe tenera</i>	<i>Gloioxanthomyces nitidus</i>	<i>Lycoperdon pyriforme</i>	<i>Stereum sanguinolentum</i>
<i>Coprinopsis atramentaria</i>	<i>Gomphidius borealis</i>	<i>Marasmiellus perforans</i>	<i>Suillus americanus</i>
<i>Coprinus comatus</i>	<i>Gymnopilus penetrans</i>	<i>Marasmius androsaceus</i>	<i>Suillus glandulosus</i>
<i>Cortinarius acutus</i>	<i>Gymnopus alpinus</i>	<b><i>Marasmius rotula</i></b>	<i>Suillus granulatus</i>
<i>Cortinarius alboviolaceus</i>	<i>Gymnopus dryophilus</i>	<i>Marasmius wittensteinii</i>	<i>Suillus neoalbidipes</i>
<i>Cortinarius armillatus</i>	<b><i>Gymnopus lachnophyllus</i></b>	<i>Melanoleuca brevipes</i>	<i>Suillus placidus</i>
<i>Cortinarius bolaris</i>	<i>Gymnosporangium cornutum</i>	<i>Merismodes fasciculata</i>	<i>Sutorius eximius</i>
<i>Cortinarius caperatus</i>	<b><i>Gymnosporangium nidus-avis</i></b>	<i>Mucronella calva</i>	<i>Taphrina robinsoniana</i>
<i>Cortinarius cinnamomeus</i>	<i>Hapalopilus rutilans</i>	<i>Mycena rubromarginata</i>	<i>Tilachlidium brachiatum</i>
	<i>Helvella lacunosa</i>	<i>Mycena sanguinolenta</i>	<i>Trametes versicolor</i>
	<i>Hemileccinum subglabripes</i>	<i>Myxarium nucleatum</i>	<i>Tremella foliacea</i>
	<i>Humaria hemisphaerica</i>	<b><i>Nectriopsis violacea</i></b>	<i>Trichaptum abietinum</i>
	<i>Hydnum repandum</i>	<i>Oxyporus populinus</i>	<i>Trichaptum laricinum</i>
	<i>Hydnum umbilicatum</i>	<i>Panaeolina foenicicii</i>	<i>Tricholoma transmutedans</i>
	<i>Hygrocybe acutoconica</i>	<i>Panellus stipticus</i>	<i>Turbinellus floccosus</i>
	<i>Hygrocybe cantharellus</i>	<i>Panellus violaceofulvus</i>	<i>Tylopilus felleus</i>
	<i>Hygrocybe chlorophana</i>	<i>Paxillus involutus</i>	<i>Tyromyces chioneus</i>
	<i>Hygrocybe conica</i>	<i>Peziza badia</i>	<i>Xerocomus subtomentosus</i>
	<i>Hygrocybe miniata</i>	<b><i>Peziza emilia</i></b>	<i>Xeromphalina enigmatica</i>

# What do These Data Mean?

Andrus Voitk



Moving our foray to a month before peak mushroom season produced an obvious expectation:

The 2017 species count would be lower than past years.

Michael and the board were very concerned that a lack of mushrooms might make the experience unsatisfactory for participants, but on three occasions I encouraged him to go ahead, because there are always fungi everywhere, adding two more expectations:

The lack of “normal” mushrooms would promote the collection of “6-O” species, (odd, obscure, or otherwise ordinarily-overlooked),

and, more importantly,

We would capture early-season species, missed by past (later-season) forays.

Then followed an unusually long, hot and dry summer, raising another expectation:

The species count would be lower still, particularly with respect to early-season fleshy mushrooms.

These matters were foremost in the organizers’ minds as foray time neared, and trails were scouted almost daily to find productive ones. Some selected

trails had to be abandoned (one ordinarily good trail had only one old polypore along its entire length!) with last minute substitutions, and there was more trepidation in the air around organizers than I have seen before. All the worry was wasted: despite the challenges, this foray had enthusiasm and excitement almost on par with our very early forays,



*Tilachlidium brachiatum*. Several collections of this strange white-needled eater of old agaric fruiting bodies were found. Perhaps a future issue of *OMF* should be devoted to such mycophagous fungi.



when nobody knew what to expect and everything was a fresh experience. All participants spoke to seemed to enjoy themselves thoroughly.

What does our dist tell us of how these expectations turned out? The title banner shows the graph of our cumulative dist (the rising red line) and the number of species per foray (the green columns along the bottom). The total of 224 species for 2017 (including three Myxomycete species, CB17, last column to the right) is, as was suspected, low for our forays. Only five regular forays have had less, but in those forays contribution by a faculty foray was non-existent or insignificant.

So, yes, not surprisingly, expectations 1 & 4 came true: holding the foray a month before peak mushroom season in a year with an unusually dry summer did have its effect. At the same time, the result was not a calamity: over 224 species is still quite a respectable number. Participants seemed to be very satisfied, and identifiers could work at a civilized pace, taking time to identify most collections.

We added 39 new species to our cumulative dist, 17% of the identified species, which is in keeping with the proportion in recent years. Intuitively you may suspect that the proportion of new species in a region could be expected to decrease slowly, as the years roll on. However, expectation 2 suggested that the proportion of new species might rise in a year with less to find, because more 6-O species tend to be brought in. Apparently not so, at least this time.

This is a bit subjective, but I noted 14 species that might be viewed as 6-O members (some collected in past years as well), or 6%. We have had more in other, much more productive years. Still, these make for a fascinating view of the diversity of fungi, and for that reason some are used to add colour to this discussion. A few other unusual finds are treated separately elsewhere.

One of the main reasons for an earlier date is to recover early-season species that we normally miss. Scanning the dist, nothing really strikes the eye as a particularly early species. Specifically, there are a lot of russulas that fruit before our normal forays. We had very few russulas, with only one species new to the dist. Also, we had very few species of *Entoloma* and *Lactarius*, and no *Clitocybe* or *Infundibulicybe*, all genera containing some early season species. Amanitas, many also early risers, were present,



*Nectriopsis violacea*. A very cool fungus, with a different diet: one of few known Myxomycete eaters, in its case, *Fuligo septica*. For a long time I thought purple was a phase *F. septica* went through, but, no—that was the contribution of *N. violacea*, eating it. Awes you with its inventive lifestyle, if not its beauty.

but no new species were seen. Possibly we have collected all the species of the genus in the province. *Inocybe* was the other early genus that stood out: seven species, four of them new. Thus, expectation 3 did not come to pass, as predicted by expectation 4, which did: absence of early season fleshy fungi was likely a direct reflection of the hot summer. Two significant exceptions of interest to the mycophagist. Two (and the less common two, at that) of our three known chanterelle species were recovered—this but months after their presence was first described. Secondly, many collections of *Boletus edulis* were



*Phaeoheliotium epiphyllum*. A small yellow cup fungus with a white stem and underside, growing, as its name suggests, on leaves. Its healthy leafy diet readily separates it from *Bisporella citrina*, *Pithiya vulgaris*, *Lachnellula agassizii*, *Byssonectria terrestris*, and many other similar little yellow cups, none of which we collected this time.



*Boletus edulis*. Collected often during the foray, and really became prominent after—a bumper year for this species in our area.

brought in, another delicious early-season species. This was probably one of the better years for that species, and as soon as a bit of rain came (after the foray, of course), the forests seemed to swell with them, even in areas where we had not encountered them before.

I cannot leave consideration of timing without commenting on the presence of Henry Beker, renowned student of the genus *Hebeloma*. It, along with *Tricoloma*, *Hygrophorus*, and a few others, is a late season genus here, often incompletely surveyed because our normal forays are too early. Of the three genera mentioned, only one collection of one species of *Tricholoma* was brought in. Not a single *Hebeloma* for Henry. However, on several occasions he graciously pointed out that he had been advised of this before coming, and was quite content.

Even with our low numbers, not all that was brought in was identified. This never happens in any year. Identifiers select from the tables material that they know, that interests them, or that otherwise appeals to them. Only later, if there is time, do they pick up strange specimens that require a lot of work to identify. Not efficient to spend three hours with one specimen, letting hundreds back up. Every once in a while some of us does a “table-clearing blitz”, trying to put a name on everything we know immediately, just to get it out of the way. This helps clear the tables, but generally removes relatively common species that have already been recorded several times, so does not directly help capture the richness.

Fine, now we know that some collections do not get identified, for a variety of reasons. But how well do we do with the material we do identify? According to the raw data, seven identified taxa did not have a specimen kept (or the specimen had to be discarded because it was improperly dried), a loss of about 3%, and less, if you count the specimens identified to genus, but not species, which are excluded from the “Species list” by definition. Not bad.



# Survey Of The Lichen-Forming Ascomycetes Collected During The 2017 NL Foray

Chris Deduke, André Arsenault and Michele Piercey-Normore

This year there were 282 lichen specimens collected from 11 trails, which included 102 species. Some of the highlights of the lichen foray this year are the findings of 40 species (and subspecies) of the genus *Cladonia*, 13 species of lichens that contain cyanobacteria as a photobiont, and three species of pin lichens. New records for the foray include *Biatora vernalis*, *Calicium lenticulare*, *Cladonia bacilliformis*, *Cladonia cyanipes*, *Cladonia furcata*, *Cladonia pocillum*, *Polyblastia hyperborea*, *Romjularia lurida*, *Solorina saccata*, *Thelotrema lepadinum* and *Vulpicida juniperina*.

Three notable species that are rarely collected in the province include *Polyblastia hyperborea*, *Romjularia lurida* and *Vulpicida juniperina*. *Polyblastia hyperborea* has previously been collected from Newfoundland and Labrador (Thomson 1997) and is likely to be



Figure 1. *Vulpicida juniperina* growing among rocks. Photo by André Arsenault.

fairly common. However, it may be undercollected because it appears as small black bumps on rocks and is easily overlooked. It was collected by Waghorne in 1896 at L'Anse Au Mort (currently known as L'Anse Amour) according to records in the CNALH (2017). *Romjularia lurida* was collected on soil over rock which appeared to be calcium-enriched. It was first identified to be *Psora globifera* but the identification was later corrected to be *R. lurida* by Chris Lewis. They look very similar in appearance but *R. lurida* has an apothecial margin which is darker than the hymenial surface, whereas the apothecial margin is lighter than the hymenium in *P. globifera* (Nash et al. 2007). *Vulpicida juniperina* (syn. *Vulpicida tilesii*) is previously known from the calcium rich soils in NL (Pitcher 2011). On Table Mountain in this year's foray, it was very common on the ground (Fig. 1). It is a yolk yellow foliose lichen which grows flat over calcareous soil in large patches and does not have soredia or rarely apothecia whereas the other common species in NL, *V. pinastri*, almost always produces abundant yellow soredia along the margins. While this species was locally

common on Table Mountain, it has rarely been reported in southwestern NL on calcium enriched soil (Pitcher 2011).

The number of species for each trail did not necessarily reflect the lichens present in those trails for various reasons including the time given to collecting lichens, the type of habitat, and the interests of the participants. This meant that the collections from some trails were few in number because lichens were not the focus of the trails. On the other hand, both Table Mountain and Barachois Pond Provincial Park showed a good representation of the lichens for those trails and they contained the highest number of species, 43 species each.

Table Mountain was an elevated exposure with calcareous soil and rocks. The top of the mountain was exposed to the northwest and was covered in sporadic tuckamore becoming a spruce forest further to the southeast (Fig. 2). It had *Cetraria aculeata*, *Flavocetraria nivalis*, *Ochrolechia androgyna*, *O. frigida*, *Pertusaria macounii*, *P. consocians*, *P. plittiana*, and *Solorina saccata*. It was also the location in which *Romjularia lurida* and *Vulpicida juniperina* were collected. Table

Mountain also contained some calcium rich habitat which explains the presence of several calcium-loving species such as *C. cariosa* and *C. pocillum*, but it also had plant debris and rocks substrates that were more acidic.



Figure 2. Southeastern view of Table Mountain with open rock barrens, tuckamore and spruce forests in the background. Photo by André Arsenault.

Barachois Pond Provincial Park was a mixed wood forest with spruce, white birch, yellow birch, balsam fir, alder and eastern white pine (Fig. 3). Its boreal nature with a more ameliorated habitat was represented by more typical boreal lichen species such as *Alectoria sarmentosa*, *Cladonia subulata*, *C. verticillata*, *Loxospora ochrophaea*, five species of *Peltigera*, *Ramalina dilacerata*, and *Usnea dasopoga*.



Figure 3. Interior forest of Barachois Pond Provincial Park. Photo by Chris Deduke.

The Stephenville bog is a seaside bog with the influence of salt spray from the nearby ocean (Fig. 4). The bog contains *Sphagnum* and other mosses, pitcher plants and sundews, but had a margin of spruce-larch forest. This bog contained some species which tend to be more typical of bogs, such as *Cladonia maxima*, *C. stygia*, and others that also have a tendency to be present in harsher conditions such as *C. cyanipes* and *Ramalina roeslerii*.



Figure 4. View of Stephenville Bog. Photo by André Arsenault.

Some species are generalists and were collected along many of the trails. One of these includes *Alectoria sarmentosa*, which is yellow pendant lichen that is often mistaken for an *Usnea* species. *Alectoria* has the appearance of white scratches on a straw coloured surface whereas *Usnea* often has a bumpy surface but is always without the scratch markings. *Alectoria* species also lack the elastic central cord which is diagnostic of *Usnea* species.

The *Cladonia chlorophaea* species complex is a complex of different cup-forming species that are often difficult to distinguish without determining the chemical contents (chemospecies) present in the thallus (Culberson *et al.* 1985). *Cladonia chlorophaea sensu stricto* is the most common species within the complex of species but *C. merochlorophaea* is a chemospecies present in NL and tends to be found on acidic substrates in moist habitats such as bogs. *Cladonia ochrochlora* is a powder horn lichen commonly found on decaying wood but can be present on other substrates also. It is difficult to distinguish from *C. coniocraea* but can be distinguished as described in Schram *et al.* (2013). *Hypogymnia physodes* (Fig. 5) is a hollow epiphyte with a white upper surface, a black under surface and upturned lobes with powdery sore-

dia. It is common on many conifers and is sometimes found mixed with *H. tubulosa*, which has narrow lobe tips with powdery soredia on the upper surface of the lobe tips. *H. physodes* can also be found on the ground in exposed locations. *Platismatia glauca* is the rag lichen with a large thin thallus that looks shredded along the edges and is common on many conifers. It has a smooth white upper surface and a black lower surface. Another species of *Platismatia*, *P. norvegica* (Fig. 6), was also collected during the foray. *P. norvegica* is a similar size but the upper surface has ridges with depressions between them and the tips are often browned (Brodo *et al.* 2001).



Figures 5 & 6. *Hypogymnia physodes* on larch (left). *Platismatia norvegica* on larch (right). Photos by André Arsenault.

While the eleven trails of the 2017 foray underwent varying intensities of collection effort, 102 species were collected in total and 11 were new species to the foray. A number of noteworthy collections were made, and the collections provided the opportunity to clarify the similarities between a number of common morphologically similar species.

We thank Chris Lewis for confirming the identification of *Romularia lurida* and we acknowledge the many participants who collected lichens that contributed to this list.

## References

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# Lichen Species Collected During Foray 2017

	Pipeline-Gorge Trail	Blowmedown Ski Club	Pasadena Ski Trail	Pasadena Watershed	Barachois Pond P. Park	Table Mountain	Stephenville Bog	Blowmedown P. Park	Barry's Lookout	Massey Drive	North Shore-Wild Cove
<i>Alectoria sarmentosa</i>	•	•	•	•	•						
<i>Amandinea punctata</i>					•						
<i>Baeomyces rufus</i>				•							
<i>Biatora vernalis</i>						•					
<i>Bryoria fremontii</i>			•								
<i>Calicium lenticulare</i>					•						
<i>Cetraria aculeata</i>						•					
<i>Cetraria islandica</i>						•					
<i>Chaenotheca chrysocephala</i>					•						
<i>Chaenotheca furfuracea</i>					•						
<i>Cladonia amaurocraea</i>						•					
<i>Cladonia arbuscula</i>					•	•	•				
<i>Cladonia bacilliformis</i>						•					
<i>Cladonia boryi</i>					•						
<i>Cladonia botrytes</i>						•					
<i>Cladonia caespiticia</i>					•						
<i>Cladonia cariosa</i>						•					
<i>Cladonia cenotea</i>					•		•				
<i>Cladonia chlorophaea complex</i>	•	•	•		•	•	•	•			
<i>Cladonia coniocraea</i>			•						•		
<i>Cladonia cornuta</i>					•						
<i>Cladonia crispata</i>					•						
<i>Cladonia cristatella</i>		•		•	•	•		•			
<i>Cladonia cyanipes</i>							•				
<i>Cladonia deformis</i>		•									
<i>Cladonia digitata</i>					•						
<i>Cladonia fimbriata</i>			•								
<i>Cladonia furcata</i>						•					
<i>Cladonia gracilis ssp. elongata</i>						•					
<i>Cladonia gracilis ssp. gracilis</i>						•					
<i>Cladonia gracilis ssp. turbinata</i>		•	•	•							
<i>Cladonia macilenta</i>					•	•					
<i>Cladonia maxima</i>		•					•				



	Pipeline-Gorge Trail	Blowmedown Ski Club	Pasadena Ski Trail	Pasadena Watershed	Barachois Pond P. Park	Table Mountain	Stephenville Bog	Blowmedown P. Park	Barry's Lookout	Massey Drive	North Shore-Wild Cove
<i>Cladonia multiformis</i>						•					
<i>Cladonia ochrochlora</i>	•			•	•	•					
<i>Cladonia pleurota</i>			•	•	•			•			
<i>Cladonia pocillum</i>						•					
<i>Cladonia pyxidata</i>						•					
<i>Cladonia rangiferina</i>			•	•	•		•	•			
<i>Cladonia rei</i>	•									•	
<i>Cladonia scabriuscula</i>					•	•		•			
<i>Cladonia squamosa</i>	•				•			•		•	
<i>Cladonia stellaris</i>							•				
<i>Cladonia stygia</i>					•	•	•				
<i>Cladonia subulata</i>					•						
<i>Cladonia symphycarpa</i>						•					
<i>Cladonia turgida</i>								•			
<i>Cladonia uncialis</i>						•					
<i>Cladonia verticillata</i>					•						
<i>Cladonia wainioi</i>									•		
<i>Dibaeis baeomyces</i>		•									
<i>Flavocetraria nivalis</i>						•					
<i>Graphis scripta</i>			•	•	•		•	•			•
<i>Hypogymnia physodes</i>	•		•		•	•	•				
<i>Hypogymnia tubulosa</i>							•				
<i>Icmadophila ericetorum</i>			•	•	•						
<i>Imshaugia aleurites</i>					•						
<i>Lecanora symmicta</i>						•					
<i>Lecidella stigmatea</i>						•					
<i>Lepraria finkii</i>	•		•		•	•		•			
<i>Lobaria pulmonaria</i>	•			•							
<i>Lobaria quercizans</i>	•										
<i>Lobaria scrobiculata</i>	•										
<i>Lopadium disciforme</i>			•								
<i>Loxospora ochrophaea</i>					•			•			
<i>Melanelixia subaurifera</i>					•						
<i>Melanohalea septentrionalis</i>							•				
<i>Mycoblastus sanguinarius</i>	•		•				•				