

ANNOTATED CUMULATIVE SPECIES LIST 2003-2012 ¹

2003-2012: 1,417 species of fungi (including 144 species of lichenized ascomycetes), plus 17 species of slime molds.

Compiled by Michael Burzynski, Andrus Voitk, Mycological consultant: Dave Malloch

[Help using this list](#)

The list is primarily Friesian (classified by morphology), with a very modest nod to phylogenetic relationships, arranging species alphabetically within the major groups. To help you find what you are looking for, here are the major, primarily Friesian, groupings:

BASIDIOMYCETES

Gilled mushrooms

Light spores

Agaricales

Russulales

Pink spores

Brown spores

Dark spores

Boletes

Tooth Fungi

Club & Coral Fungi

Puffballs, Stinkhorns & False Truffles

Polypores

Jelly Fungi

Tough basidiomycetes with smooth to veined spore bearing surface

Rusts, Smuts & other phytoparasitic basidiomycetes

ASCOMYCETES

Lichenized Ascomycetes

Operculate Discomycetes

Inoperculate Discomycetes

Pyrenomycetes

Hemiascomycetes

Plectomycetes

Anamorphs

Zygomycetes

SLIME MOLDS

BASIDIOMYCETES

Gilled mushrooms

Light coloured (white) spores
(Leucosporus)

Agaricales

Amanita albocreata
*Amanita bisporigera*²
*Amanita ceciliae*²²
Amanita elongata
Amanita flavoconia
Amanita fulva
*Amanita groenlandica*²⁰
Amanita mortenii
Amanita muscaria
Amanita nivalis
Amanita porphyria
Amanita rubescens
Amanita sinicoflava
Amanita sp. "GRL02"¹⁹
Amanita sp. "Killdevil amanita"³
Amanita sp. "NFL02"¹⁹
Amanita sp. "NFL03"¹⁹
Amanita sp. "NFL04"¹⁹
Amanita sp. "NFL05"¹⁹
Amanita sp. "NFL06"¹⁹
Amanita sp. "NFL09"¹⁹
Amanita sp. "NFL10"¹⁹
Amanita sp. "NFL11"¹⁹
Amanita sp. "precocious amanita"⁷
Amanita vaginata var. *alba*²³
Amanita vaginata var. *vaginata*
Amanita wellsii
Ampulloclitocybe clavipes
Armillaria ostoyae
Armillaria sinapina
Arrhenia acerosa
Arrhenia acerosa f. *latispora*
Arrhenia griseopallida
Arrhenia obatra
Arrhenia obscurata
Arrhenia onisca
Arrhenia philonotis
Arrhenia retiruga
Arrhenia rustica
Arrhenia sphagnicola
Arrhenia velutipes
Asterophora parasitica
Blasiphalia pseudogrisella
Callistosporium luteoolivaceum
Cantharellula umbonata

Cantharellus roseocanus
Catathelasma imperiale
Catathelasma ventricosum
Cheimonophyllum candidissimum
Clitocybe candicans
Clitocybe candicans var. *dryadicola*
Clitocybe dealbata
Clitocybe deceptiva
Clitocybe diatretra
Clitocybe festiva
Clitocybe foetens
Clitocybe globispora
Clitocybe lateritia
Clitocybe metachroa
Clitocybe sinopica
*Clitocybe subalpina*²⁰
Clitocybe vibecina
Clitocybula familia
Collybia cirrhata
Collybia cookei
Collybia tuberosa
Connopus acervatus
Craterellus lutescens
Craterellus tubaeformis
Crinipellis setipes
*Cystoderma amianthinum*¹⁵
Cystoderma fallax
*Cystoderma jasonis*¹⁵
Cystodermella cinnabarinus
Cystodermella granulosa
Fayodia anthracobia
Gymnopus alpina
Gymnopus confluens
Gymnopus dryophilus
Gymnopus impudicus
Gymnopus loiseleurietorum
Gymnopus obscurus
Gymnopus ocior
Gymnopus peronatus
Gyroflexus brevbasiatus
Hemimycena gracilis
Hemimycena lactea
Hohenbuehelia petaloides
Hohenbuehelia reniformis
Hohenbuehelia tremula
Humidicutis marginata var. *marginata*
Humidicutis marginata var. *olivacea*
Humidicutis pura
Hygrocybe acutoconica
Hygrocybe borealis^{32, 47}
Hygrocybe cantharellus
Hygrocybe ceracea

Hygrocybe chlorophana
Hygrocybe cinerella
Hygrocybe coccinea
Hygrocybe coccineocrenata
Hygrocybe colemanniana
Hygrocybe conica
Hygrocybe conica var. *chloroides*
Hygrocybe conica var. *conicopalustris*
Hygrocybe helobia
Hygrocybe insipida
*Hygrocybe irrigata*²⁵
*Hygrocybe lacmus*³²
Hygrocybe laeta
*Hygrocybe lilacina*³²
Hygrocybe miniata
Hygrocybe miniata var. *mollis*
Hygrocybe mycenoides
Hygrocybe nitida
Hygrocybe phaeococcinea
*Hygrocybe pratensis*³²
Hygrocybe psittacina
Hygrocybe punicea
Hygrocybe reidii
Hygrocybe singeri var. *albifolia*
Hygrocybe squamulosa
Hygrocybe substrangulata var. *rhodophylla*
Hygrocybe turunda
Hygrocybe turunda var. *sphagnophila*
Hygrophoropsis aurantiaca
Hygrophoropsis morganii
Hygrophoropsis rufa
Hygrophorus agathosmus
Hygrophorus camarophyllus
Hygrophorus chrysodon
Hygrophorus discoideus
Hygrophorus eburneus
Hygrophorus erubescens
Hygrophorus gliocyclus
Hygrophorus inocybiformis
Hygrophorus monticola
Hygrophorus olivaceoalbus
Hygrophorus piceae
Hygrophorus pudorinus
Hygrophorus purpurascens
Hygrophorus russula
Hygrophorus speciosus
Hypsizygos ulmarius
Infundibulicybe gibba
*Infundibulicybe gigas*³⁸
Infundibulicybe squamulosa
*Laccaria altaica*²⁷

*Laccaria bicolor*²⁷
*Laccaria laccata*²⁷
*Laccaria longipes*²⁷
Laccaria nobilis
*Laccaria proxima*²⁷
*Laccaria striatula*²⁷
*Laccaria tortilis*²⁷
*Laccaria trullisata*²⁷
Lentinellus micheneri
Lepiota cristata
Lepiota cortinarius
Lepista gilva
Lepista graveolens
Lepista martiorum
Lepista multiformis
Lepista nuda
Lepista pseudectypa
Leucoagaricus leucothites
Lichenomphalia alpina
Lichenomphalia hudsoniana
Lichenomphalia umbellifera
Limacella illinita
Lyophyllum connatum
Lyophyllum decastes
Lyophyllum fuligineum
Lyophyllum fumosum
Lyophyllum semitale
Marasmiellus perforans
Marasmiellus vaillantii
Marasmius androsaceus
Marasmius capillaris
Marasmius epiphyllus
Marasmius graminum
Marasmius oreades
Marasmius pallidocephalus
Marasmius thujinus
*Megacollybia rodmanii*³⁷
Melanoleuca alboflavida
Melanoleuca cognata
Melanoleuca melaleuca
Mycena abramsii
*Mycena adonis*¹⁴
Mycena alexandri
Mycena atralboides
Mycena borealis
Mycena capillaripes
Mycena cinerella
Mycena clavicularis
Mycena citrinomarginata
Mycena diosma
Mycena epipterygia
Mycena epipterygioides
Mycena filopes
Mycena flavoalba
Mycena floridula
Mycena galericulata
Mycena galopoda
Mycena haematopus
Mycena hemisphaerica
Mycena laevigata
Mycena latifolia
Mycena leptocephala
Mycena maculata
Mycena megaspora
Mycena metata
Mycena murina
Mycena oregonensis
Mycena picta
Mycena pura
Mycena pura var. *alba*
Mycena rorida
Mycena rosea
Mycena rosella
Mycena rosella var. *albida*
Mycena rubromarginata
Mycena sanguinolenta
Mycena septentrionalis
Mycena strobilinoidea
Mycena stylobates
Mycena urania
Mycena vulgaris
Mycena zephira
Mycenella trachyspora
Mycetinis scorodonius
Omphalina pyxidata
Panellus ringens
Panellus stipticus
Panellus violaceofulvus
Phyllostopsis nidulans
Pleurocybella porrigens
Pleurotus dryinus
Polyozellus multiplex
Pseudoomphalina pachyphylla
Rhizomarasmius epidryas
Rhodocollybia butyracea var. *butyracea*
Rhodocollybia prolixa var. *distorta*
Rhodocollybia maculata
Rhodocollybia maculata var. *scorzonerea*
Rickenella fibula
Rickenella mellea
*Rugosomyces carneus*³³
Rugosomyces fallax
Sarcomyxa serotina
Tectella patellaris
Tephrocybe palustris
Tephrocybe stripilea
Tricholoma albobrunneum
Tricholoma apium
Tricholoma arvernense
Tricholoma atrodiscum
Tricholoma atosquamosum
Tricholoma caligatum
Tricholoma columbetta
Tricholoma davisiae
Tricholoma dulciolens
Tricholoma equestre
Tricholoma flavobrunneum
Tricholoma flavum
Tricholoma focale
*Tricholoma fulvum*⁴²
Tricholoma fumosoluteum
Tricholoma hemisulphureum
Tricholoma imbricatum
Tricholoma inamoenum
Tricholoma inodermeum
Tricholoma intermedium
Tricholoma leucophyllum
Tricholoma luridum
Tricholoma magnivelare
Tricholoma myomyces
Tricholoma pardinum
*Tricholoma pessundatum*⁴²
Tricholoma portentosum
Tricholoma roseoacerbum
Tricholoma saponaceum
Tricholoma scalpturatum
Tricholoma serratifolium
Tricholoma sp. "Rusty trich"
Tricholoma sp. "Unearthly trich"
Tricholoma stans
Tricholoma striparophyllum
*Tricholoma subsejunctum*⁴⁴
Tricholoma sulphureum
Tricholoma terreum
*Tricholoma transmutans*⁴²
Tricholoma ustale
Tricholoma vaccinum
Tricholoma virgatum
Tricholoma viridilutescens
Tricholomopsis decora
Tricholomopsis rutilans
Tricholomopsis sulfureoides
Xeromphalina campanella
Xeromphalina caudicinalis
Xeromphalina cornui
Xeromphalina fellea

Russulales

Lactarius affinis var. *affinis*

<i>Lactarius affinis</i> var. <i>viridilactis</i>	<i>Lactarius vellereus</i>	<i>Russula rhodopoda</i>
<i>Lactarius alpinus</i>	<i>Lactarius vietus</i>	<i>Russula rosacea</i>
<i>Lactarius aquizonatus</i>	<i>Lactarius vinaceorufescens</i>	<i>Russula rosea</i>
<i>Lactarius aurantiacus</i>	<i>Lactarius zonarius</i>	<i>Russula roseipes</i>
<i>Lactarius badiusanguineus</i>	<i>Lactarius zonaroides</i>	<i>Russula sanguinea</i>
<i>Lactarius blumii</i>	<i>Russula abietina</i>	<i>Russula silvicola</i>
<i>Lactarius camphoratus</i>	<i>Russula adusta</i>	<i>Russula sphagnophila</i>
<i>Lactarius chrysorrhoeus</i>	<i>Russula aeruginea</i>	<i>Russula variata</i>
<i>Lactarius circellatus</i>	<i>Russula albonigra</i>	<i>Russula velenovskyi</i>
<i>Lactarius deceptivus</i>	<i>Russula altaica</i>	<i>Russula versicolor</i>
<i>Lactarius deliciosus</i> ¹²	<i>Russula aquosa</i>	<i>Russula vesca</i>
<i>Lactarius deterrimus</i> ¹²	<i>Russula barlae</i>	<i>Russula violaceoincarnata</i>
<i>Lactarius fumosus</i>	<i>Russula betularum</i>	<i>Russula viscida</i>
<i>Lactarius gerardii</i>	<i>Russula betulina</i>	<i>Russula xerampelina</i>
<i>Lactarius glyciosmus</i>	<i>Russula brevipes</i>	
<i>Lactarius griseus</i>	<i>Russula brunneola</i>	<u>Pink spores (Hyporhodium)</u> ³¹
<i>Lactarius helvus</i>	<i>Russula chamiteae</i>	<i>Clitopilus prunulus</i>
<i>Lactarius hibbardae</i>	<i>Russula chloroides</i>	<i>Clitopilus scyphoides</i> var.
<i>Lactarius hysginus</i> var. <i>hysginus</i>	<i>Russula cicatricata</i>	<i>omphaliformis</i>
<i>Lactarius leonis</i>	<i>Russula citrinochlora</i>	<i>Clitopilus scyphoides</i> var.
<i>Lactarius lignyotus</i> var.	<i>Russula claroflava</i>	<i>scyphoides</i> f. <i>reductus</i>
<i>canadensis</i> ³⁹	<i>Russula clavipes</i>	<i>Entoloma abortivum</i>
<i>Lactarius lignyotus</i> var. <i>lignyotus</i> ³⁹	<i>Russula compacta</i>	<i>Entoloma alpicola</i>
<i>Lactarius lignyotus</i> var.	<i>Russula consobrina</i>	<i>Entoloma bicolor</i>
<i>marginatus</i> ³⁹	<i>Russula cyanoxantha</i>	<i>Entoloma bloxamii</i>
<i>Lactarius lignyotus</i> var.	<i>Russula decolorans</i>	<i>Entoloma caeruleopolitum</i>
<i>nigroviolascens</i> ³⁹	<i>Russula delica</i>	<i>Entoloma carbonicola</i>
<i>Lactarius mucidus</i>	<i>Russula densifolia</i>	<i>Entoloma cetratum</i>
<i>Lactarius nitidus</i>	<i>Russula dissimulans</i>	<i>Entoloma cuspidiferum</i>
<i>Lactarius oculatus</i>	<i>Russula emetica</i>	<i>Entoloma elodes</i>
<i>Lactarius peckii</i>	<i>Russula exalbicans</i>	<i>Entoloma fuscomarginatum</i>
<i>Lactarius pseudoflexuosus</i>	<i>Russula firmula</i>	<i>Entoloma fuscotomentosum</i>
<i>Lactarius pseudouvidus</i>	<i>Russula foetens</i>	<i>Entoloma infula</i>
<i>Lactarius pubescens</i>	<i>Russula fragilis</i>	<i>Entoloma lagenicystis</i>
<i>Lactarius representaneus</i>	<i>Russula gracillima</i>	<i>Entoloma lepiotosmus</i>
<i>Lactarius resimus</i>	<i>Russula graminea</i>	<i>Entoloma lividocyanulum</i>
<i>Lactarius rosezonatus</i>	<i>Russula heterophylla</i>	<i>Entoloma longistriatum</i>
<i>Lactarius rufus</i>	<i>Russula laurocerasi</i>	<i>Entoloma luridum</i>
<i>Lactarius salicis-herbaceae</i>	<i>Russula mairei</i>	<i>Entoloma majaloides</i>
<i>Lactarius salicis-reticulatae</i>	<i>Russula nana</i>	<i>Entoloma pallescens</i>
<i>Lactarius scrobiculatus</i> var.	<i>Russula nauseosa</i>	<i>Entoloma papillatum</i>
<i>canadensis</i>	<i>Russula nigricans</i>	<i>Entoloma prunuloides</i>
<i>Lactarius sordidus</i> ⁴⁶	<i>Russula nitida</i>	<i>Entoloma quadratum</i>
<i>Lactarius sphagneti</i>	<i>Russula norvegica</i>	<i>Entoloma rhodopolium</i> var.
<i>Lactarius subdulcis</i>	<i>Russula ochroleuca</i>	<i>nidorosum</i>
<i>Lactarius subvellereus</i>	<i>Russula ochroleuroides</i>	<i>Entoloma rhodopolium</i> var.
<i>Lactarius tabidus</i>	<i>Russula olivacea</i>	<i>rhodopolium</i>
<i>Lactarius theiogalus</i>	<i>Russula paludosa</i>	<i>Entoloma sericatum</i>
<i>Lactarius thynios</i> ¹²	<i>Russula peckii</i>	<i>Entoloma sericellum</i>
<i>Lactarius torminosulus</i>	<i>Russula praeumbonata</i>	<i>Entoloma sericeum</i>
<i>Lactarius torminosus</i>	<i>Russula puellaris</i>	<i>Entoloma sinuatum</i>
<i>Lactarius trivialis</i>	<i>Russula purpurata</i>	<i>Entoloma sp. "dune entoloma"</i> ⁵
<i>Lactarius tuomikoskii</i>	<i>Russula queletii</i>	<i>Entoloma strictius</i>
<i>Lactarius uvidus</i>	<i>Russula raoultii</i>	

Entoloma strictius var. *isabellinum*
Entoloma strictum
Entoloma subsepiaceum
Entoloma turbidum
Entoloma xanthoserrulatum
*Pluteus cervinus*³⁵
Pluteus atomarginatus
Pluteus salicinus
Rhodocybe caelata
Rhodocybe hirneola
Rhodocybe nitellina

Brown spores (Cortinaria +
Derminus)

Agrocybe erebia
Agrocybe paludosa
Agrocybe pediades
Alnicola geraniolens
Alnicola melinoides
Alnicola sphagneti
Alnicola tantilla
Bolbitius titubans
Conocybe filaris
Conocybe fimetaria
Conocybe lactea
Conocybe pilosella
Conocybe semiglobata
Conocybe tenera
Conocybe watlingii
Cortinarius acutus
Cortinarius agathosmus
Cortinarius alborufescens
Cortinarius albovariegatus
Cortinarius alboviolaceus
Cortinarius alnetorum
Cortinarius alpinus
Cortinarius americanus
Cortinarius angelesianus
Cortinarius anomalellus
Cortinarius anomalus
Cortinarius armeniacus
Cortinarius armillatus
Cortinarius atrocaeruleus
Cortinarius aureofulvus
Cortinarius badiovinaceus
Cortinarius balteatus
Cortinarius bataillei
Cortinarius bibulus
Cortinarius biformis
Cortinarius bivelus
Cortinarius bolaris
Cortinarius brunneus var. *brunneus*
Cortinarius callisteus

Cortinarius camphoratus
Cortinarius caninus
Cortinarius caperatus
Cortinarius casimiri
Cortinarius chrysolithus
Cortinarius cinnabarius
Cortinarius cinnamomeus
Cortinarius claricolor
Cortinarius clarobrunneus
Cortinarius collinitus
Cortinarius colus
Cortinarius corrugis
Cortinarius crassus
Cortinarius croceofolius
Cortinarius croceus
Cortinarius decipiens
Cortinarius delibutus
Cortinarius depressus
Cortinarius diasemospermus var.
leptospermus
Cortinarius disjungendus
Cortinarius erubescens
Cortinarius evernius
Cortinarius favrei
Cortinarius fennoscandicus
Coertinarius fervidus
Cortinarius flexipes var. *flabellus*
Cortinarius flexipes var. *flexipes*
Cortinarius flexipes var. *inolens*
Cortinarius flos-paludis
Cortinarius fragrans
Cortinarius fulvescens
Cortinarius fulvo-ochraceus
Cortinarius furvolaesus
Cortinarius gentilis
Cortinarius glandicolor
Cortinarius glaucopus
Cortinarius grosmorensis
Cortinarius helvelloides
Cortinarius hemitrichus
Cortinarius hercynicus
Cortinarius heterocyclus
Cortinarius hinnuleus f. *subtypicus*
Cortinarius huronensis
Cortinarius illuminus
Cortinarius imbutus
Cortinarius impennis
Cortinarius incisus
Cortinarius incognitus
Cortinarius infractus
Cortinarius ionophyllus
Cortinarius junghuhnii
Cortinarius laetissimus
Cortinarius laniger

Cortinarius leucophanes
Cortinarius lilacinus
Cortinarius limonius
Cortinarius lucorum
Cortinarius luteo-ornatus
Cortinarius malachius
Cortinarius malicorius
Cortinarius mucifluus
Cortinarius mucosus
Cortinarius multiformis
Cortinarius obtusus
Cortinarius ochrophyllus
Cortinarius olivaceofuscus
Cortinarius paleaceus
Cortinarius paragaudis
Cortinarius paragaudis ssp.
oenochelis
Cortinarius parvannulatus
Cortinarius percomis
Cortinarius pholideus
Cortinarius pilatii
Cortinarius pluvius
Cortinarius polaris
Cortinarius porphyropus
Cortinarius prestigiosus
Cortinarius privignoides
Cortinarius purpurascens
Cortinarius quarcticus
Cortinarius raphanoides
Cortinarius renidens
Cortinarius rubellus
Cortinarius rusticus
Cortinarius saginus
Cortinarius sanguineus
Cortinarius saturninus
Cortinarius scaurus
Cortinarius scaurus var.
sphagnophilus
Cortinarius scutulatus
Cortinarius semisanguineus
Cortinarius semivestitus
Cortinarius septentrionalis
Cortinarius sertipes
Cortinarius solis-occasus
Cortinarius sommerfeltii
Cortinarius sphagnophilus
Cortinarius stemmatus
*Cortinarius stillatitius*⁴⁸
Cortinarius subcroceofolius
Cortinarius suberi
Cortinarius subtortus
Cortinarius tortuosus
*Cortinarius traganus*³⁴
Cortinarius triformis

Cortinarius trivialis
Cortinarius tubarius
Cortinarius turmalis
Cortinarius uliginosus
Cortinarius umbrinolens
Cortinarius valgus
Cortinarius varius
Cortinarius venustus
Cortinarius vespertinus
Cortinarius vibratilis
Cortinarius violaceus
Crepidotus mollis
Crepidotus versutus
Galerina atkinsoniana
Galerina calyptrata
Galerina hypnorum
Galerina leptocystis
*Galerina marginata*⁹
Galerina mniophila
Galerina paludosa
Galerina sphagnicola
Galerina sphagnorum
Galerina stagnina
Galerina tibiicystis
Galerina uncialis
Galerina vittaeformis var.
vittaeformis f. *tetraspora*
Gymnopilus bellulus
Gymnopilus junonius
Gymnopilus penetrans
Gymnopilus picreus
Gymnopilus sapineus
Hebeloma bruschetii
Hebeloma cavipes
Hebeloma crustuliniforme
Hebeloma helodes
Hebeloma hiemale
Hebeloma incarnatum
Hebeloma lutense
Hebeloma mesophaeum
Hebeloma nigellum
Hebeloma polare
Hebeloma sinapizans
Hebeloma strophosum
Hebeloma vaccinum
Hebeloma velutipes
Inocybe asterospora
Inocybe calamistrata
Inocybe castanea
Inocybe dulcamara
Inocybe fastigiata
Inocybe flocculosa
Inocybe fraudans
Inocybe fraudans var.

chamaesalicis
Inocybe fuscomarginata
Inocybe geophylla var. *geophylla*
Inocybe geophylla var. *lilacina*²⁶
Inocybe gymnocarpa
Inocybe lacera
Inocybe lanuginosa
Inocybe leucoblema
Inocybe leptophylla
Inocybe longispora
Inocybe microspora
Inocybe mixtilis
Inocybe nappies
Inocybe perbrevis
Inocybe petiginosa
Inocybe rennyi
Inocybe soluta
Inocybe stellatospora
Inocybe subcarpta
Inocybe teraturga
Inocybe umboninota
Inocybe virgata
Kuehneromyces lignicola
Kuehneromyces mutabilis
*Paxillus involutus*³⁰
*Paxillus rubicundulus*³⁰
*Paxillus vernalis*³⁰
Phaeocollybia gregaria
Phaeocollybia jennyae
Pholiota alnicola
Pholiota astragalina
Pholiota highlandensis
Pholiota lenta
*Pholiota limonella*⁴¹
Pholiota mixta
Pholiota scamba
Pholiota spumosa
Pholiota squarrosa
Pholiota squarrosoides
Pholiotina arrhenii
Ripartites tricholoma
Simocybe centunculus
Simocybe reducta
Tubaria confragosa
Tubaria furfuracea
Tubaria minutalis

Dark (purple-brown to black)
spores (Pratella)

Agaricus bitorquis
Agaricus campestris
Agaricus haemorrhoidarius
Agaricus micromegethus

*Agaricus silvicola*⁴⁵
Chroogomphus ochraceus
Coprinopsis atramentaria
Coprinopsis ephemeroides
Coprinopsis epichloea
Coprinopsis nivea
Coprinopsis parvula
Coprinopsis stercorea
Coprinus comatus
*Gomphidius borealis*⁴⁹
*Gomphidius glutinosus*⁴⁹
*Gomphidius maculatus*⁴⁹
*Gomphidius subroseus*⁴⁹
Hypholoma capnoides
Hypholoma dispersum
*Hypholoma elongatum*¹¹
Hypholoma ericaeum
Hypholoma eximium
Hypholoma fasciculare
Hypholoma marginatum
Hypholoma myosotis
Hypholoma radicosum
Hypholoma subericaceum
Hypholoma sublateralium
Hypholoma udum
Panaeolus acuminatus
Panaeolus campanulatus
Panaeolus foenicicii
Panaeolus semiovatus
Panaeolus sphinctrinus
Panaeolus subbalteatus
Parasola plicatilis
Psathyrella candolleana
Psathyrella conissans
Psathyrella lutensis
Psathyrella piluliformis
Psathyrella sarcocephala
Psathyrella septentrionalis
Psathyrella sphagnicola
Psathyrella velutina
Psilocybe coprophila
Psilocybe montana
Psilocybe phyllogena
Psilocybe semilanceata
*Stropharia alcis*¹⁶
*Stropharia arctica*¹⁶
Stropharia ambigua
Stropharia hornemanii
Stropharia magnivelaris

Boletes

Austroboletus gracilis
Boletus edulis

Boletus calopus
Boletus huronensis
Boletus subglabripes
Chalciporus piperatus
Chalciporus pseudorubinellus
*Leccinum holopus*²⁸
Leccinum rotundifoliae
Leccinum scabrum
Leccinum schistophilum
*Leccinum snellii*²⁹
Leccinum variicolor
*Leccinum versipelle*²¹
*Leccinum vulpinum*²¹
Suillus americanus
Suillus cavipes
*Suillus clintonianus*¹⁷
Suillus flavidus
Suillus glandulosus
Suillus granulatus
*Suillus grevillei*¹⁷
Suillus grisellus
Suillus intermedius
Suillus luteus
Suillus neoalbidipes
Suillus paluster
Suillus placidus
*Suillus serotinus*⁸
Suillus spectabilis
Suillus spraguei
Suillus subalutaceus
Suillus umbonatus
Tylopilus chromapes
Tylopilus eximius
Tylopilus felleus
Tylopilus porphyrosporus
Xanthoconium affine
Xerocomus badius
*Xerocomus gracilis*¹⁸

Hydnoid or tooth fungi

Bankera fuligneo-alba
Bankera violascens
Hericium coralloides
Hydnellum aurantiacum
Hydnellum caeruleum
Hydnellum concrescens
Hydnellum frondosum
Hydnellum geogenium
Hydnellum multiceps
*Hydnellum peckii*¹⁰
*Hydnellum pineticola*⁴³
Hydnellum scrobiculatum
Hydnellum suaveolens

Hydnellum velutinum var.
spongiosipes
Hydnum albomagnum
Hydnum repandum
Hydnum rufescens
Hydnum umbilicatum
Phellodon melaleucus
Phellodon niger var. *alboniger*
Phellodon niger var. *niger*
Phellodon tomentosus
Sarcodon glaucopus
Sarcodon imbricatus
Sarcodon leucopus
Sarcodon scabrosus
Sarcodon stereosarcinon
Sarcodon subfelleus
Sistotrema confluens
Steccherinum ochraceum

Club and coral fungi

Alloclavaria purpurea
Clavaria argillacea
Clavaria argillacea var.
sphagnicola
Clavaria falcata
Clavaria fumosa
Clavaria rosea
*Clavariadelphus ligula*⁴⁰
*Clavariadelphus sachalinensis*⁴⁰
Clavariadelphus truncatus
Clavulina cinerea
Clavulina coralloides
Clavulina rugosa
Clavulinopsis fusiformis
Clavulinopsis laeticolor
Gomphus clavatus
Gomphus floccosus
Lentaria micheneri
Mucronella calva
Multiclavula mucida
Multiclavula vernalis
Ramaria acrisiccescens
Ramaria aurantiisiccescens
Ramaria bataillei
Ramaria cartilaginea
Ramaria cystidiophora var.
cystidiophora
Ramaria cystidiophora var.
fabiolens
Ramaria fasciculata
Ramaria fennica
Ramaria flaccida
Ramaria flava

Ramaria formosa
Ramaria grundii
Ramaria harrisonii
Ramaria leptiformosa
Ramaria myceliosa
Ramaria obtusissima
Ramaria pallida
Ramaria pallidosaponaria
Ramaria rubrievanescens
Ramaria sandaracina
Ramaria stricta
Ramaria testaceoflava
Ramaria velocimutans
Ramaria virescens
Ramaria zeppelinospora
Ramariopsis crocea
Ramariopsis kunzei
Ramariopsis rufipes
Ramularia taraxaci
Trichoglossum hirsutum

Puffballs, stinkhorns and false truffles (Gasteromycetes)

Alpova cinnamomea
Bovista pila
Bovista plumbea
Calvatia cretacea
Crucibulum laeve
Dictyophora duplicata
Lycoperdon caudatum
Lycoperdon curtisii
Lycoperdon foetidum
Lycoperdon molle
Lycoperdon nigrescens
Lycoperdon ovatum
Lycoperdon pedicellatum
Lycoperdon perlatum
Lycoperdon pusillum
Lycoperdon umbrinum
Morganella pyriformis
Mutinus ravenelii
Nidularia deformis
Rhizopogon evadens
Rhizopogon pseudoroseolus
Scleroderma bovista
Sphaerobolus stellatus

Polypores

Albatrellus caeruleoporus
Albatrellus peckianus
Anomoporia myceliosa
Antrodia heteromorpha

Antrodia serialis
Antrodia variiformis
Antrodiella romellii
Basidioradulum radula
Bjerkandera adusta
Boletopsis grisea
Ceraceomyces borealis
Cerrena unicolor
Cinereomyces lindbladii
Climacocystis borealis
Coltricia perennis
Datronia mollis
Datronia scutellata
Diplomitoporus crustulinus
Diplomitoporus lenis
Fomes fomentarius
Fomitopsis ochracea
Fomitopsis pinicola
Fomitopsis rosea
Ganoderma applanatum
Gloeophyllum sepiarium
Gloeoporus taxicola
Hapalopilus nidulans
Hymenochaete cinnamomea
Hymenochaete fuliginosa
Inonotus cuticularis
Inonotus glomeratus
Inonotus obliquus
Inonotus radiatus
Ischnoderma benzoinum
Ischnoderma resinosum
Jahnoporus hirtus
Leptoporus mollis
Megalocystidium leucoxanthum
Meruliopsis taxicola
Oligoporus balsameus
Oligoporus guttulatus
Onnia circinata
Onnia tomentosa
Otidea onotica
Oxyporus populinus
Perenniporia subacida
Phaeolus schweinitzii
Phellinus chrysoloma
Phellinus cinereus
Phellinus conchatus
Phellinus contiguus
Phellinus ferreus
Phellinus ferruginosus
Phellinus gilvus
Phellinus igniarius
Phellinus laevigatus
Phellinus nigricans
Phellinus nigrolimitatus

Phellinus pini
Phellinus prunicola
Phellinus tremulae
Piptoporus betulinus
Polyporus badius
Polyporus brumalis
Polyporus lepideus
*Polyporus varius*⁶
Postia balsamea
Postia caesia
Postia fragilis
Postia ptychogaster
Postia sericeomollis
Postia stiptica
Postia tephroleuca
Ptychogaster rubescens
Skeletocutis amorpha
Trametes hirsuta
Trametes ochracea
Trametes pubescens
Trametes versicolor
*Trichaptum abietinum*²⁴
Trichaptum biforme
Trichaptum fuscoviolaceum
Trichaptum laricinum
Trichaptum subchartaceum
Tubulicrinis glebulosus
Tyromyces chioneus
Veluticeps abietina

Jelly fungi

Auricularia americana
Calocera cornea
Calocera viscosa
Dacrymyces chrysospermus
Dacrymyces lacrymalis
Dacrymyces tortus
Exidia glandulosa
Exidia saccharina
*Femsjonia peziziformis*⁵¹
Guepiniopsis alpina
Pseudohydnum gelatinosum
Tremella encephala
Tremella foliacea
Tremella mesenterica
Tremella mycetophila
Tremiscus helvelloides

Tough basidiomycetes with a smooth to veined spore-bearing surface

Aleurodiscus amorphus

Aleurodiscus laurentianus
Aleurodiscus penicillatus
Alutaceodontia alutacea
Amphinema byssoides
Amylocorticium subsulphureum
Amylostereum chailletii
Athelia decipiens
Athelia epiphylla
Athelopsis subinconspicua
Basidiodendron caesiocinereum
Boidinia furfuracea
Boidinia propinqua
Botryobasidium conspersum
Botryobasidium isabellinus
Botryobasidium medium
Botryobasidium subcoronatum
Botryobasidium vagum
Byssocorticium pulchrum
Ceraceomyces eludens
Ceraceomyces microsporus
Chondrostereum purpureum
Coniophora arida
Coniophora olivacea
Coniophora puteana
*Cotylidia undulata*⁵⁰
Cylindrobasidium evolvens
Cytidia salicina
Dacryobolus sudans
Globulicium hiemale
Gloeocystidiellum leucoxanthum
Gloeocystidiellum porosum
Gloeodontia subasperispora
Gloiothele citrina
Gloiothele lactescens
Henningsomyces candidus
Hymenochaete rubiginosa
Hymenochaete tabacina
Hymenoscyphus lutescens
Hyphoderma argillaceum
Hyphoderma sambuci
Hyphoderma setigerum
Hyphoderma sibiricum
Hyphodontia alutaria
Hyphodontia arguta
Hyphodontia aspera
Hyphodontia breviseta
Hyphodontia crustosa
Hyphodontia hastata
Hyphodontia pallidula
Hyphodontia quercina
Hyphodontia rimosissima
Hyphodontia spathulata
Hyphodontia subalutacea
Hydnophlebia chrysorhiza

Laurilia sulcata
Leifia flabelliradiata
Leucogyrophana lichenicola
Leucogyrophana romellii
Merismodes anomala
Merismodes fasciculata
Mycoacia uda
Paulliticium allantosporum
Peniophora aurantiaca
Peniophora erikssonii
Peniophorella praetermissa
Peniophorella pubera
Perenniporia medulla-panis
Perenniporia subacida
Perenniporia variegata
Phanerochaete sanguinea
Phanerochaete sordida
Phanerochaete velutina
Phlebia livida
Phlebia tremellosa
Phlebiella christiansenii
Phlebiopsis gigantea
Piloderma bicolor
Piloderma croceum
Piloderma fallax
Plicatura nivea
Plicaturopsis crispa
Pseudotomentella tristis
Resinicium bicolor
Scytinostroma galactinum
Scytinostroma odoratum
Sistotrema octosporum
Sistotremastrum suecicum
Skeletocutis lenis
Skeletocutis vulgaris
Stereum hirsutum
Stereum ochraceoflavum
Stereum rugosum
Stereum sanguinolentum
Stromatocyphella conglobata
Stypella subgelatinosa
Stypella vermiformis
Thelephora palmata
Thelephora terrestris
Tomentella botryoides
Tomentella fuscocinerea
Tomentella lapida
Tomentella lateritia
Tomentella radiosa
Tomentella stiposa
Tomentella sublilacina
Tomentella terrestris
Trechispora confinis
Vararia gallica

Vararia investiens
Vararia racemosa
Vesiculomyces citrinus
Xenasmatella vaga

Rusts, smuts and other phytoparasitic basidiomycetes

Anthracoidea heterospora
Anthracoidea kariii
Anthracoidea laxae
Chrysomyxa ledi
Chrysomyxa rhododendri
Cystobasidium hypogymniicola
Exobasidium cassandrae
Exobasidium juelianum
Exobasidium karstenii
Exobasidium ledi
Exobasidium oxycocci
Exobasidium rhododendri
Exobasidium savilei
Exobasidium splendidum
Exobasidium vaccinii
Exobasidium vaccinii-uliginosi
Gymnosporangium cornutum
Melampsora caryophyllacearum
Melampsora epitea
Melampsora populina
Melampsoridium betulinum
Naohidemycetes vacciniorum
Nyssopsora clavellosa
Phragmidium rubi-idaei
Puccinia conglomerata
Puccinia dioicae
Puccinia fergussonii
Puccinia poarum
Puccinia polygoni
Puccinia ribis
Pucciniastrum arcticum
Pucciniastrum epilobii
Pucciniastrum goeppertianum
Pucciniastrum potentillae
Uredinopsis americana

ASCOMYCETES

Lichenized ascomycetes

Alectoria sarmentosa
Arctopermelia centrifuga
Baeomyces rufus
Bellemerea cinereorufescens
Bryoria americana

Bryoria furcellata
Bryoria fuscescens
Bryoria trichodes
Buellia punctata
Buellia punctata
Candellariella aurella
Cetraria islandica ssp. crispiformis
Cetraria muricata
Cladonia albonigra
Cladonia arbuscula
Cladonia arbuscula ssp. squarrosa
Cladonia boryi
Cladonia caespiticia
Cladonia carneola
Cladonia cenotea
Cladonia chlorophaea
Cladonia cornuta
Cladonia crispata
Cladonia cristatella
Cladonia deformis
Cladonia digitata
Cladonia farinacea
Cladonia gracilis ssp. gracilis
Cladonia gracilis ssp. turbinata
Cladonia gracilis ssp. elongata
Cladonia grayi
Cladonia groenlandica
Cladonia macilenta
Cladonia maxima
Cladonia metacorallifera
Cladonia mitis
Cladonia multififormis
Cladonia ochrochlora
Cladonia phyllophora
Cladonia pleurota
Cladonia pyxidata
Cladonia rangiferina
Cladonia rei
Cladonia scabriuscula
Cladonia squamosa
Cladonia stellaris
Cladonia strepsilis
Cladonia stygia
Cladonia subulata
Cladonia sulphurina
Cladonia symphycarpa
Cladonia terrae-novae
Cladonia turbinata
Cladonia turgida
Cladonia uncialis
Cladonia verticillata
Degelia plumbea
Dibaeis baeomyces
Diploschistes scruposus

Ephebe lanata
Flavocetraria nivalis
Gowardia nigricans
Haematomma elatinum
Hypogymnia incurvoides
Hypogymnia physodes
Hypogymnia tubulosa
Hypogymnia vittata
Icmadophila ericetorum
Imshaugia aleurites
Japewia subaurifera
Lecanora allophana
Lecanora intricata
Lecanora polytropa
Lecanora symmicta
Lecidea lulensis
Lecidea pullata
Leptogium saturninum
Lobaria pulmonaria
Lobaria quercizans
Loxospora ochrophaea
Melanelia hepatizon
Melanelixia subaurifera
Melanohalea trabeculata
Mycoblastus affinis
Mycoblastus sanguinarius
Mycocalicium subtile
Nephroma arcticum
Nephroma bellum
Nephroma laevigatum
Ochrolechia androgyna
Ochrolechia frigida
Parmelia saxatilis
Parmelia squarrosa
Parmeliella triptophylla
Parmeliopsis ambigua
Parmeliopsis capitata
Parmeliopsis hyperopta
Peltigera aphthosa
Peltigera canina
Peltigera hymenina
Peltigera leucophlebia
Peltigera malacea
Peltigera neopolydactyla
Peltigera polydactylon
Peltigera praetextata
Peltigera rufescens
Pertusaria amara
Pertusaria dactylina
Phaeophyscia pussiloides
Placynthiella uliginosa
Platismatia glauca
Platismatia norvegica
Porpidea flavocoerulescens

Protoparmelia badia
Radulomyces confluens
Radulomyces hiemalis
Ramalina dilacerata
Ramalina roesleri
Ramalina thrausta
Ramboldia cinnabarina
Rhizocarpon geographicum
Staurothele fissa
Stereocaulon alpinum
Stereocaulon condensatum
Stereocaulon dactylophyllum
Stereocaulon paschale
Stereocaulon saxatile
Stereocaulon tomentosum
Stereocaulon vesuvianum
Trapeliopsis granulosa
Trechispora farinacea
Trechispora microspora
Tubulicrinis calothrix
Tubulicrinis gracillimus
Tuckermanopsis americana
Tuckermanopsis sepincola
Umbilicaria muhlenbergii
Umbilicaria polyphylla
Usnea dasypoga
Usnea filipendula
Usnea longissima
Varicellaria rhodocarpa
Vulpicida pinastri
Xylographa abietina
Xylographa vitiligio

Operculate discomycetes

Aleuria aurantia
Ascobolus ciliatus
Ascobolus stercorarius
Cheilymenia fimicola
Cheilymenia stercorea
Coprotus luteus
Dermea cerasi
Geopyxis carbonaria
Helvella corium
Helvella crispa
Helvella elastica
Helvella lacunosa
Helvella macropus
Helvella phlebophora
Helvella sulcata
Humaria hemisphaerica
Iodophanus carneus
Melastiza chateri
Neottiella vivida

Octospora rubens
Orbilia curvatispora
Otidea onotica
Peziza alcis
Peziza arvernensis
Peziza badia
Peziza brunnea
Peziza domiciliana
Peziza praetervisa
Peziza repanda
Peziza succosa
Peziza tenacella
Rhizina undulata
Scutellinia cejpui
Scutellinia heterosculcurata
Scutellinia kerguelensis
Scutellinia nigrohirtula
Scutellinia scutellata
Scutellinia superba
Tapesia hydrophila

Inoperculate discomycetes

Ascocoryne cylichnium
Ascocoryne sarcoides
Ascocoryne turficola
Bisporella citrina
Bryoglossum gracile
Bryoglossum rehmi
*Chlorociboria aeruginascens*⁴
*Chlorociboria aeruginosa*⁴
Cistella acuum
Coryne dubia
Crociceras coronatum
Cudonia circinans
Cudonia confusa
Cyathicula coronata
Dasyscyphus virgineus
Encoelia furfuracea
Erysiphe sordida
Geoglossum cookeianum
Geoglossum glabrum
Geoglossum umbratile
Helotium clavus
Heyderia abietis
Hyaloscypha albohyalina
Hymenoscyphus calyculus
Hymenoscyphus imberbis
Hymenoscyphus lutescens
Hymenoscyphus perilis
Hymenoscyphus scutula
Lachnellula agassizii
Lachnellula suecica
*Leotia lubrica*¹³

*Leotia viscosa*¹³

Microglossum rufum

Mollisia cinerea

Neocudoniella radicea

Neolecta irregularis

Podophacidium xanthomelum

Rhytisma ilicis-canadensis

Rhytisma prini

Rhytisma salicinum

Rutstroemia firma

Spathularia flavida

Spathulariopsis velutipes

Thuemenidium arenarium

Pyrenomycetes

Apiosporina morbosa

Claviceps purpurea

Diatrype bullata

Diatrype stigma

Elaphocordyceps ophioglossoides

Helminthosphaeria clavariarum

Hypocrea alutacea

Hypocrea pulvinata

Hypomyces chrysospermus

Hypomyces hyalinus

Hypomyces lactifluorum

Hypomyces lateritius

*Hypomyces leotiicola*¹³

Hypomyces luteovirens

Hypomyces microspermus

Hypomyces torminosus

Hypomyces viridilutescens

Hypoxyton fuscum

Melanoconis silbestoma

Nectriopsis candicans

Plectocarpon lichenum

Hemiascomycetes

Taphrina betulina

*Taphrina robinsoniana*³⁶

Plectomycetes

Onygena equina

Anamorphs

Cladosporium herbarum

Pseudocercospora leptosperma

Ramularia destructiva

Zygomycetes

Endogone pisiformis

Spinellus fusiger

SLIME MOULDS

Arcyria incarnata

Ceratomyxa fruticulosa

Didymium clavus

Didymium iridis

Didymium melanospermum

Didymium minus

Didymium squamulosum

Fuligo cinerea

Fuligo septica

Leocarpus fragilis

Lycogala epidendrum

Physarum confertum

Physarum virescens

Physarum viride

Stemonitis axifera

Stemonitis fusca

Tubifera ferruginosa

1. We do not consider identifications done under foray conditions incontrovertible. All are open to a variable, small but real margin of error. They are also subject to changes brought about by new information. These notations attest to that. In many cases identifiers indicate some degree of uncertainty by the use of qualifiers such as *Lactarius sordidus.*, cf., coll., complex, group, sl. and the like. These qualifiers are retained in the raw data and the original cards are kept, all as evidence about the identifier's assessment of certainty. However, these qualifiers are omitted on the published list for ease of reading. Our raw data is available for review. This list may err by commission or omission. Where there is a choice, our preference has been to err on the side of omission, ie we prefer to leave out a species that may be here over claiming one exists that does not live here.
2. *Amanita bisporigera*. Includes specimens identified as *Amanita virosa*. According to *Amanita* expert Rod Tulloss, one of our guest faculty for four forays, DNA testing has revealed that *Amanita virosa* is an European species that does not fruit here. Rod is of the opinion that the same species has both a two-spored and four-spored form, depending on the season and its stage of maturity.
3. *Amanita* sp. ("Killdevil amanita"). Identified by Rod Tulloss as a hitherto undescribed species, initially given the code name "NFL07", but provisionally renamed by him after Killdevil Lodge, where our forays took place when this species was found. Discussed and illustrated in Voitek: *A little illustrated book of common mushrooms of Newfoundland and Labrador*.
4. Microscopic distinction between *C. aeruginascens* and *C. aeruginosa* made first in 2008. Earlier reports may have contained both under one name.
5. Hitherto undescribed species, according to Machiel Noordeloos. "Dune entoloma" will be changed to proper binomial if studies confirm Machiel's opinion and he formally describes it with new name. We look forward to using its new name soon; meanwhile we refer to it as "*Entoloma* sp." or use the nickname, given because it was found in sand dunes.
6. Our concept of *P. varius* is that it encompasses *P. elegans* and is synonymous with *P. leptocephalus*.
7. *Amanita* sp. ("precocious amanita") corresponds to *Amanita* sp32 of Tulloss, thought to be undescribed. Discussed and illustrated in
 - a. Despres et al.: *Mille et un champignons du Québec*. (CD-ROM).
 - b. Lamoureux: *Champignons du Québec, Tome 2, Les Amanites*.
 - c. Tulloss: URL < <http://pluto.njcc.com/~ret/amanita/mainaman.html>>
 - d. McNeil: *Le grand livre des champignons du Québec et de l'est du Canada*
 - e. Voitek: *A little illustrated book of common mushrooms of Newfoundland and Labrador*
 In the first three references, above, it is given a provisional name "Amanita praecox Lamoureux", under which name it appears on some North American foray lists and recent books. Such provisional names for undescribed spp are declared invalid by Article 34.1 of the International Code of Botanical Nomenclature and are therefore not accepted by most taxonomists. We hope that this and other similarly treated species will be described by their discoverers in a timely fashion, so that the whole world can enjoy use of these new names for these new species without feeling conscience stricken as a sinner against good taxonomic practice.
8. We have not been entirely convinced that we were have found more than one species of slimy larch *Fuscoboletinus*, whether pale greenish grey (which often has been called *F. aeruginascens* elsewhere) or deep russet brown (which certainly fits *F. serotinus*). Since *F. aeruginascens* = *F. viscidus* = *F. laricinus* (all based on European types), and *F. serotinus* is a North American name for a taxon in the same complex, we have followed the approach of *Le cercle des mycologues de Montréal* by considering all four taxa synonymous until further work is done to elucidate their interrelationship, if any. We shall keep track of the original identifications, so that should the species be considered separate, the identifiers' determination can be noted.
9. Because DNA has shown them to be synonymous, species identified as *Galerina autumnalis* have been changed to *G. marginata*; this epithet covers both identifications.
10. *Hydnellum peckii* and *H. diabolus*. All our collections came from the same trails in 2003, 2004, 2005. They were identified as *H. peckii* in 2003 and 2005 but in 2003 were identified as *H. diabolus* by an identifier, who felt the two were not synonymous. Because we have at no time identi-

fied both, to show we can differentiate between them, because they are considered synonymous by *Index Fungorum* and many other authors and because the majority of identifiers side with *H. peckii*, we have arbitrarily reclassified them as such. With this comment we acknowledge the possibility that they may be distinct and our *H. peckii* may contain some or all *H. diabolus*. Until better clarification, we shall keep them as is for the sake of ease, since all but one identifier is in agreement, but shall be glad to reclassify them in the face of better evidence than we have at present.

11. *Hypholoma elonagatum* contains specimens identified as *H. elonagatipes* because they are synonymized in *Nordic Macromycetes*.

12. According to Jorinde Nuytinck, *Lactarius deliciosus* is a European species that is not found here. *L. deterrimus* is also a European species; erroneously one or more similar species have been called *L. deterrimus* or *L. deliciosus* var. *deterrimus* in North America. In 2007 Jorinde did genetic studies on all collections from my (av) private collection, collected all over the island. Only two species were identified: *L. thyinos* and *L. "deterrimus"*. These were the only species identified 2006 and 2007. 2003–2005 forays identified *L. deliciosus*; although incorrect, this is allowed to stand, as we lack sufficient evidence to make accurate retrospective corrections. *L. "deterrimus"* has not been renamed because work is still being done to see whether this represents one or more species. Until then we'll use the epithet, full well knowing it to be incorrect. For the sake of ease of reading, we have elected to omit the quotation marks and retain the italics in the list, but note the problems here.

13. 2003–2006 we have only recorded *Leotia lubrica* and *L. viscosa*, often including with the former mushrooms with some olive to greenish tone on cap ± stem. Some authors have considered all in-between shades as *L. atrovirens*, while others reserve this epithet only for mushrooms that are evenly dusky green. We intended to examine this in closer detail in 2007. However, at that time we learned of the report by Rogerson and Samuels, suggesting the dusky green variety may be a parasite, *Hypomyces leotiicola*, on *L. lubrica*. We did identify such parasitized fungi in 2007 and recorded the parasite. Further reading after the foray suggests that there may also be at least one, if not more, species of all-green leotias, unrelated to any

parasite. They are described as having gel in the outer layer of the stem. Since we did not examine for that among our foray material, we are unable to make an identification of *L. atrovirens* for 2007 or earlier.

14. Identifications of *Mycena amabilissima* have been included with *M. adonis*. Although not all authors synonymize the two, we have been unable to define meaningful differences between the two identifications. We suspect this is correct but accept that we may hide two species under one name and are prepared to reconsider, if new evidence comes to light.

15. *Cystoderma amianthinum* and *C. jasonis* are two macroscopically similar species having amyloid basidiospores. According to *Nordic Macromycetes* *C. amianthinum* should have ellipsoidal to oblong spores and white flesh while *C. jasonis* should have more or less fusoid basidiospores and yellow to ochre flesh. In addition *C. jasonis* should have "arthrospores" in the upper part of the flesh of the pileus. Material from Newfoundland and New Brunswick are not very consistent for the first two of these characters and never have the arthrospores. Further study is needed, including examination of "authentic" *C. jasonis* from Europe.

16. All identifications of *Stropharia semiglobata* from 2003 have been renamed *S. alcis*. Once aware of the difference in 2004, we checked spore size of our collections: all fit with *S. alcis*. Subsequently we have checked a few specimens randomly and based on this, have found only *S. alcis* in our woods—until 2010, when another "routine" check uncovered one *S. arctica* and four *S. alcis*. Finding *S. arctica* on the Great Northern Peninsula and not elsewhere (so far) suggests this is a northern species (as its name suggests) and we may encounter more there as well as in Labrador. The specimen was young and grew on mouse dung. Although its stem was considerably shorter than that of *S. alcis*, in proportion to the cap diameter, that may be due to immaturity, not a morphologic feature of this species.

17. Initially we considered the dark red-brown and the yellow version as synonyms of *Suillus grevillei*. In 2007 we were shown that these have been officially separated as distinct species in Scandinavia, reserving *S. grevillei* for the yellow version, common there, as originally described. The dark red-brown version is known as *S. clintonianus*,

as described by Peck for the version most commonly encountered in North America. Since this is a North American name and since there is at least geographic, if not ecologic, difference between the two, we elected to separate them into those two species in 2007. An attempt to “correct” our records retroactively met with the usual difficulty: not every collection of this very common mushroom was photographed or dried. Therefore, we elected to let identifications before 2007 stand as made, with this note as qualifier. A rough estimate is that about one-third of specimens brought in were yellow (*S. grevillei*). This is higher than their relative distribution in our woods but it seems foragers are more prone to bring in a more unusual specimen. Parenthetically, it is my (av) guess that DNA will eventually show them to be synonymous and that the difference is due to a minor typographic error in the gene controlling red-brown pigment. I suspect that the yellow version (flavino?) is no more a different species than an albino in other organisms. However, until this has been shown or disproven, since there is a North American name for a North American mushroom, it seems more correct taxonomically to treat them as separate. Lumping later is no problem.

18. All identifications of *Boletus/Boletellus/Xerocomus* tomentosus/subtomentosus have been changed to *X. gracilis*. For his research, Andy Taylor asked for specimens of these species. Specimens of all our foray material as well as all specimens in my (av) private collections, collected all over the island and the southern coast of Labrador, were sent. After DNA analysis we were informed that all, no matter how different in gross morphology or geographic origin, mapped out with *X. gracilis*. On this basis it seemed most accurate to consider this our only or most frequent representative of this species complex. We acknowledge that all subsequent specimens will not undergo DNA analysis and the name may hide a close relative but think that until we are able to separate them by other means this will be the most accurate, even if not perfect, way to report them.
19. These are species identified by Rod Tulloss as hitherto undescribed. Tulloss’s initial code numerics have been used for identification. Tulloss spent four forays with us and as a result created a picture-book of *Amanita* species in Newfoundland and Labrador on his website < <http://pluto.njcc.com/~ret/amanita/mainaman.html>>. These species are undergoing more study and some may be withdrawn as new, while others may end up synonymized. Once reported in the literature, we shall uncode them and identify them accordingly. Keeping the code name in the meantime allows us to track them through this process. As seen on his web site, Tulloss feels there are even more undescribed spp. in our province.

20. This may be a misidentification. While we have confirmed (collections outside our forays) that *A. groenlandica* does indeed grow in our province, Tulloss now feels this species is not it and probably represents another member of the *A. fulva*-*A. vaginata* complex. Please see Tulloss’s website for further details.
 21. We have decided to follow the genetic studies of den Bakker et al., considering all reddish-capped leccinums growing in conifer woods as *L. vulpinum* in the broad sense (s.l.). Thus, this epithet lumps together determinations *L. piceinum* from conifer woods as well as *L. atrostipitatum*, *L. aurantiacum* and *L. versipelle* from primarily coniferous mixed woods. We recognize that den Bakker’s experience with North American leccinums is not as exhaustive as his experience with European leccinums and species like *L. piceinum* may prove to be good. Where originally identified as such, those changes can be reverted. We also recognize that *L. atrostipitatum*, *L. aurantiacum* and *L. versipelle* from primarily coniferous mixed woods may have been birch associates (a few birch being the only significant deciduous tree in these woods), but since the woods are primarily coniferous and since we have not seen reddish-capped leccinums in pure birch woods, we feel this lumping may be more accurate at this stage of our knowledge. In this, we agree with Kuo that in view of the confusion of profusion with leccinums, perhaps less is better until a clearer understanding of what we have in North America emerges. Again, should new evidence emerge, the original identifications are on record and can be changed back, as indicated.
- We did not follow the above guidelines in the case of two collections from Battle Harbour, Labrador. Both were made from barrens, with only dwarf trees. Only dwarf birch was seen around them, with no conifers nearby. These have been listed as identified, *L. versipelle*. Other collections identified

- as *L. versipelle* and *L. atrostipitatum* from Labrador, but coming from primarily coniferous mixed woods, even if a birch was noted in the vicinity, have been listed for the moment as *L. lupinum*, for the reasons given above.
22. This is the name of an European mushroom, which reputedly does not grow in North America. The species found here is closest to the European species by gross morphology. Not examined microscopically. By gross morphology, seems to fit the description of “*Amanita* V3” of Tulloss, a species as yet not formally described; Tulloss has also referred to it by a provisional name, “*Amanita borealisorora*”; further, it seems to conform to the species to which Lamoureux has given a provisional name “*Amanita rhacopus*”, also lacking a formal description to date. Again, we hope that this and other similarly treated species will be described by their discoverers in a timely fashion, so that the whole world can enjoy use of these new names for these new species without feeling conscience stricken as a sinner against good taxonomic practice.
 23. Best fit on gross morphology with *Amanita vaginata* var. *alba*, although its stem was not quite as long in proportion to cap diameter as expected and a tiniest tinge of yellow-pink on disc. That species is described by Lamoureux as “*Amanita albiceps*”, a name awaiting formal description before coming into common use. Not examined microscopically but photo and voucher specimens available for further study.
 24. Synonymous with *Hirschioporus abietinus*.
 25. *H. irrigata* synonymized with *H. unguinosa* and contains specimens identified as the latter.
 26. Synonymized with *I. lilacina* and *I. geophylla* var. *violacea*.
 27. In 2008 laccarias keyed out as special project. Vast majority keyed out as *L. bicolor*, four *L. striatula* and one *L. longipes*; no *L. laccata* or *L. proxima*. In other years field identification primary mode of determining species. We do not know whether the difference is due to difference in ecoregion, an unusual year or method of identification.
 28. Synonymized with *L. niveum*. This is the earlier name and therefore should have preference. However, there is some uncertainty about the species originally defined under that epithet. That, plus the fact that *L. holopus* is entrenched in common usage, has made us prefer the latter.
 29. In 2008, *Leccinum snellii* was closest fit macroscopically, but spore size was closer to *L. flavostipitatum*, with little overlap; did not have yellow stipe of the latter and did not dry yellow.
 30. Primarily differentiated by ecologic habitat. *P. rubicundulus* identified by growth among alders, with no other trees in neighbourhood; some morphological differences from *P. involutus* noted on most occasions. *P. vernalis* identified by growth among poplars, with no other trees nearby; little if any morphological difference from *P. involutus* noted. *P. involutus* considered an associate of birch, conifers or mixed forest.
 31. We have elected to follow the ranking of Noorde-loos regarding entolomas and have dispensed with genera *Alboleptinoa*, *Leptonia*, *Nolanea* and others, considering these and others as subgenera of Genus *Entoloma*.
 32. We are following ranking of Boertmann and considering all species in *Camarophyllus* as *Hygrocybe*.
 33. We have treated *C. persicolor* as a probable synonym.
 34. *C. traganus* includes specimens identified as *C. pyriodoris*.
 35. *P. cervinus* contains specimens identified as *P. atricapillus*.
 36. *Taphrina robinsoniana*—causes alder tongue gall in *Alnus rugosa*. The gall is morphologically indistinguishable from that caused in European *A. incana* (ssp. *incana*) by *T. alni*, but apparently the latter is not a North American species. We have only *A. (incana ssp.) rugosa* and *A. (viridis ssp.) crispa* on the Island; alder tongue gall is very host-specific and is a parasite of the former only.
 37. Northeastern North American relative of the European *M. platyphylla*, confirmed with nuclear sequencing by Ron Petersen.
 38. We have elected to use the name of the European equivalent, *pro tem*, until evidence appears that ours is a new species. *Clitocybe maxima*, referring to a species of temperate Europe, has been declared a *nomen dubium*. Harmaja described a similar species in northern Europe as *C. gigas*. Because of our northern location we selected the name of the northern species, until ours can be checked.
 39. We did not scrutinize our *Lactarius lignyotus* in detail, until in 2008, when unexpectedly we saw that one had turned a startling purple after several hours on the display table. Subsequent check of

specimens and past photos has showed that all members of this species that we have seen have marginate gills. This excludes *L. lignyotus* var. *canadensis*, the default variety for past collections, where variety has not been noted. All specimens collected in Newfoundland has stained pink, compatible with *L. lignyotus* var. *canadensis*. Material in Labrador has stained either pink, or after several hours a deep purple. The latter is compatible with *L. lignyotus* var. *marginatus*. Since these features have not been noted in the past, we have not changed earlier determinations. Since then, we have found a few pink-staining specimens with totally amarginate gills, keying out to *L. lignyotus* var. *lignyotus*. In 2012 we found our first purple staining specimen with amarginate gills, keying out to *L. lignyotus* var. *nigroviolascens*.

40. Before 2010, we have not checked these clavarias too closely routinely, so that pre-2010 *C. ligula* may hide *C. sachaliensis*.
41. *Pholiota aurivella* and *Pholiota limonella* are not distinguished reliably by macro- or micro-morphologic means. Since the former is a European species and the latter North American, we have elected, much as the Québec group, to arbitrarily assign the North American epithet to all ours, pending further information.
42. Until the question is resolved with genetic marker studies, we have elected to use the name *Tricholoma fulvum*. However, we recognize that this may be a purely European taxon, with

an as yet undetermined North American counterpart. Yves Lamoureux and the Québec group have suggested Peck's *Tricholoma transmutans* might fit best with our collections. Because this may well turn out to be the case, we have elected to leave identifications made to that species stand, and as Yves, consider the two concepts conspecific in our area. We also note the potential confusion between this/these species and *Tricholoma pessundatum*.

43. We follow Harrison and consider *Hydnellum pineticola* to be the North American equivalent of the European *H. ferrugineum*.
44. In Europe a conical dusky yellow-green *Tricholoma* under deciduous trees is known as *T. sejunctum* and a similar one under conifers as *T. viridilutescens*. Our only similar *Tricholoma* grows under conifers. We have elected to show either of the above two identifications as *T. subsejunctum*, a North American species, until evidence suggests a change.
45. Until evidence to suggest the contrary, we elect to consider *Agaricus abruptibulbus* as a synonym with *A. silvicola*.
46. Unpublished data from Ghent University shows that our *Lactarius sordidus* differs from the European *L. necator/L. turpis*, and best fits with *L. sordidus* Peck.
47. *Hygrocybe borealis* is the northeastern genetic sister to *H. virginea*.
48. Includes collections identified as *Cortinarius integerrimus*.
49. The identification of *Gomphidius* species has been problematic. *G. glutinosus* may contain

within it other close species, and related determinations may reflect the seeming dissatisfaction with *G. glutinosus* as a good fit. We hope to be able to clarify this soon.

50. A poor place to put this one, but since it has no gills, putting it with *Rickenella* is not much better, so here it is.
51. Phylogeny shows *Femsjonia* to clump together as a group, quite remote from *Ditiola*.